

SYMPOSIUM REVIEW

The diseasome of physical inactivity – and the role of myokines in muscle–fat cross talk

Bente K. Pedersen

The Centre of Inflammation and Metabolism at the Department of Infectious Diseases, and Copenhagen Muscle Research Centre, Rigshospitalet, the Faculty of Health Sciences, University of Copenhagen, Denmark

Type 2 diabetes, cardiovascular diseases, colon cancer, breast cancer, dementia and depression constitute a cluster of diseases, which defines ‘a diseasome of physical inactivity’. Both physical inactivity and abdominal adiposity, reflecting accumulation of visceral fat mass, are associated with the occurrence of the diseases within the diseasome. Physical inactivity appears to be an independent and strong risk factor for accumulation of visceral fat, which again is a source of systemic inflammation. Chronic inflammation is involved in the pathogenesis of insulin resistance, atherosclerosis, neurodegeneration and tumour growth. Evidence suggests that the protective effect of exercise may to some extent be ascribed to the anti-inflammatory effect of regular exercise, which can be mediated via a reduction in visceral fat mass and/or by induction of an anti-inflammatory environment with each bout of exercise. The finding that muscles produce and release myokines provides a conceptual basis to understand the mechanisms whereby exercise influences metabolism and exerts anti-inflammatory effects. According to our theory, contracting skeletal muscles release myokines, which work in a hormone-like fashion, exerting specific endocrine effects on visceral fat. Other myokines work locally within the muscle via paracrine mechanisms, exerting their effects on signalling pathways involved in fat oxidation.

(Received 28 July 2009; accepted after revision 10 September 2009; first published online 21 September 2009)

Corresponding author B. K. Pedersen: Centre of Inflammation and Metabolism, Rigshospitalet – Section 7641, Blegdamsvej 9, DK-2100, Copenhagen, Denmark. Email: bkp@rh.dk

Introduction

Most human diseases are not independent of each other, although they are often treated individually.

To understand the mechanisms underlying these networks of diseases, it is obviously necessary to map out the detailed diagram of various cellular components that are influenced by specific genes and gene products. Such genetic network-based thinking has provided insights into the pathogenesis of several diseases (Barabasi, 2007).

Another type of network in human disease is the so-called social network, which includes all human-to-human interactions (e.g. familial, friendship, sexual and proximity-based contacts) that play a role in the spread of pathogens (Barabasi, 2007). Interestingly,

social networks have an important role also in the spread of obesity. Christakis and Fowler suggest that friends have an even more important effect on a person’s risk of obesity than genes do (Christakis & Fowler, 2007). They showed that if one friend became obese during a given time interval, the other friend’s chances of following suit increased by 171%. In contrast, if one sibling became obese, the chance that the other would become obese increased by 40%. The latter study underscores the role of social networks in the obesity epidemic. In continuation, I hereby suggest the existence of a ‘lifestyle network’, or more specifically a ‘physical inactivity network’ of key importance in understanding why several diseases appear in clusters, although they are highly different in terms of phenotypical presentation and required treatment. Type 2 diabetes, cardiovascular diseases, colon cancer, postmenopausal breast cancer, dementia and depression constitute a cluster of diseases, which I hereby identify as ‘the diseasome of physical inactivity’, as appears in Fig. 1.

It is well-established that physical inactivity increases the risk of type 2 diabetes (Tuomilehto *et al.* 2001),

This review was presented at *The Journal of Physiology* Symposium on *Physiological regulation linked with physical activity and health*, which took place at the 36th International Congress of Physiological Sciences in Kyoto, Japan on 31 July 2009. It was commissioned by the Editorial Board and reflects the views of the authors.

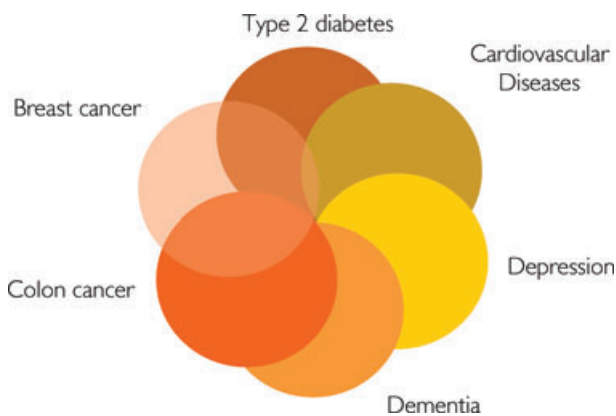


Figure 1. Type 2 diabetes, cardiovascular diseases, colon cancer, postmenopausal breast cancer, dementia and depression constitute a cluster of diseases, which can be identified as 'the disease of physical inactivity'

cardiovascular diseases (CVD) (Nocon *et al.* 2008), colon cancer (Wolin *et al.* 2009) and postmenopausal breast cancer (Monninkhof *et al.* 2007). In addition, physical inactivity may also play a role in the development of dementia (Rovio *et al.* 2005) and even depression (Paffenbarger *et al.* 1994). The latter diagnoses are not meant to represent an exclusive list of diseases or disorders related to physical inactivity. However, they are all frequent chronic diseases, associated with an enhanced risk of premature morbidity. Patients with type 2 diabetes have a markedly increased risk of CVD (Diamant & Tushuizen, 2006). In addition, type 2 diabetes is associated with impaired cognitive function as well as with both Alzheimer's disease and vascular dementia, and individuals with type 2 diabetes also have a high prevalence of affective illness, including major depression (reviewed in Komulainen *et al.* 2008). Other studies report that

type 2 diabetes is associated with an elevated risk of developing colon and breast cancer, as well as pancreatic, liver and endometrial cancer (Richardson & Pollack, 2005). There is indeed also overlap between other diagnoses within the disease of physical inactivity; however, it appears that type 2 diabetes plays a central role.

It is interesting that the disease of physical inactivity represents highly different diseases, but they share important pathogenetic mechanisms. Clearly, independently of body mass index (BMI), physical inactivity is a risk factor for all-cause mortality (Pedersen, 2007). Moreover, chronic systemic inflammation is associated with physical inactivity independent of obesity (Fischer *et al.* 2007).

I herein put forward the hypothesis that physical inactivity leads to accumulation of visceral fat and consequently the activation of a network of inflammatory pathways, which promote development of insulin resistance, atherosclerosis, neurodegeneration and tumour growth, and thereby the development of the diseases belonging to the 'disease of physical inactivity'. The hypothesis is illustrated in Fig. 2.

Physical inactivity and abdominal adiposity

A relatively large amount of subcutaneous adipose tissue has little or no damaging effect and may even offer protection against chronic diseases, whereas strong evidence exists for the detrimental effects of visceral fat and fat in the liver and in muscle (Pischon *et al.* 2008).

In this context, fat is not just fat and ectopic fat accumulation may be regarded as fat in 'the wrong places'. Abdominal adiposity is associated with cardiovascular disease (CVD) (Haffner, 2007), type 2 diabetes (Bays,

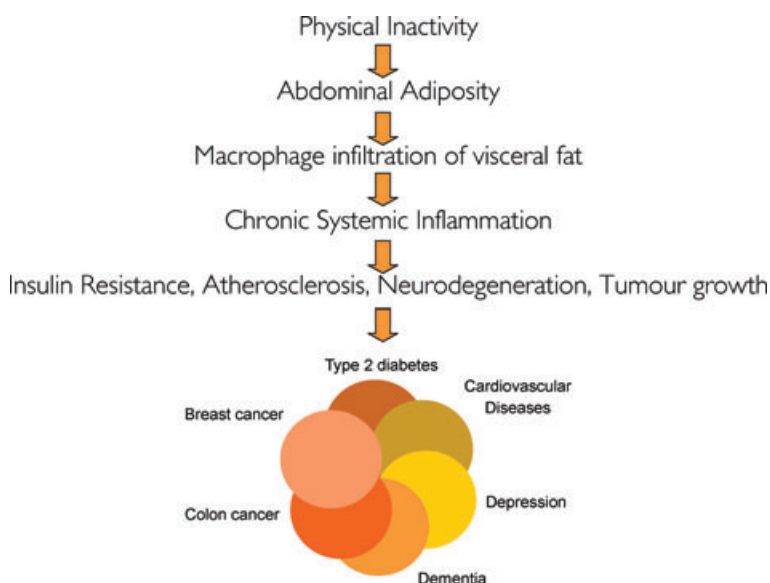


Figure 2. Hypothesis: Physical inactivity leads to accumulation of visceral fat and consequently to the activation of a network of inflammatory pathways, which promotes development of insulin resistance, atherosclerosis, neurodegeneration, and tumour growth, leading to the development of 'the disease of physical inactivity'

2009), dementia (Whitmer *et al.* 2008), colon cancer (Giovannucci, 2007) and breast cancer (Xue & Michels, 2007) as well as all-cause mortality independent of BMI, even in people with a normal body weight (Pischon *et al.* 2008). Thus, the health consequences of abdominal adiposity and physical inactivity are similar. Moreover, both physical inactivity (Pedersen & Febbraio, 2008) and abdominal adiposity (Yudkin, 2007) are associated with persistent systemic low-grade inflammation (Festa *et al.* 2002; Handschin & Spiegelman, 2008).

We recently conducted a model of physical inactivity that certainly points at a direct link between physical inactivity and accumulation of visceral fat. A group of young healthy men decreased their daily stepping for 2 weeks to 1500 steps from the range recommended for adults of around 10 000. During this time, they developed a markedly impaired glucose tolerance as well as attenuation of postprandial lipid metabolism. The intervention was associated with a 7% increase in intra-abdominal fat mass, measured by MR scanning, without a change in total fat mass while total fat-free mass and BMI decreased (Olsen *et al.* 2008). Thus, the striking consequences of physical inactivity included accumulation of visceral fat mass and impaired glucose and lipid metabolism.

Visceral fat as a cause of low-grade systemic inflammation

Models of lipodystrophy suggest that if the subcutaneous fat becomes inflamed and adipocytes undergo apoptosis/necrosis, the fat storing capacity is impaired; hence, fat is deposited as ectopic fat. Given the anti-inflammatory effects of regular exercise (Petersen & Pedersen, 2005), physical inactivity may lead to inflammation of subcutaneous adipose tissue and impaired ability to store fat also in people who do not fulfil the criteria for lipodystrophy.

Although speculative, one explanation of the differential outcome of accumulating fat subcutaneously or as ectopic fat could be that when fat is stored in 'the wrong places', it will stimulate an inflammatory response. Evidence exists that visceral fat is more inflamed than subcutaneous fat and constitutes an important source of systemic inflammation (Yudkin, 2007).

Thus, although 2 weeks of inactivity did not provoke an increase in circulating levels of cytokines, evidence exists of an association between physical inactivity and low-grade systemic inflammation in healthy, young individuals (Petersen & Pedersen, 2005). These findings would be compatible with the notion that accumulation of visceral fat precedes chronic systemic inflammation.

Inflammation as a cause of chronic diseases

Chronic inflammation promotes development of insulin resistance, atherosclerosis, neurodegeneration and tumour

growth (Handschin & Spiegelman, 2008), and thereby the development of the diseases belonging to the 'diseasome of physical inactivity'.

Mounting evidence suggests that TNF- α plays a direct role in the metabolic syndrome (recently reviewed in Pedersen & Febbraio, 2008). In short, patients with diabetes demonstrate high protein expression of TNF- α in skeletal muscle and increased TNF- α levels in plasma, and it is likely that adipose tissue, which produces TNF- α , is the main source of the circulating TNF- α . *In vitro* studies demonstrate that TNF- α has direct inhibitory effects on insulin signalling. Recently it was demonstrated that TNF- α infusion in healthy humans induces insulin resistance in skeletal muscle, without an effect on endogenous glucose production. It has also been proposed that TNF- α causes insulin resistance indirectly *in vivo* by increasing the release of free fatty acids (FFA) from adipose tissue. TNF- α increases lipolysis in human and 3T3-L1 adipocytes. However, TNF- α has no effect on muscle fatty acid oxidation, but increases fatty acid incorporation into diacylglycerol, which may be involved in the development of the TNF- α -induced insulin resistance in skeletal muscle. Moreover, evidence suggests that TNF- α plays a direct role in linking insulin resistance to vascular disease (Yudkin *et al.* 2005; Plomgaard *et al.* 2005). In cardiovascular diseases, activated immune cells also play major roles, particularly in the aetiology of atherosclerosis (Matter & Handschin, 2007).

Several downstream mediators and signalling pathways seem to provide the cross talk between inflammatory and metabolic signalling. These include the discovery of c-Jun N-terminal kinase (JNK) and I κ B kinase (I κ BK) as critical regulators of insulin action activated by TNF- α (Hotamisligil, 2003). In human TNF- α infusion studies, TNF- α increases phosphorylation of p70 S6 kinase, extracellular signal-regulated kinase-1/2 and c-Jun NH₂-terminal kinase, concomitant with increased serine and reduced tyrosine phosphorylation of insulin receptor substrate-1. These signalling effects are associated with impaired phosphorylation of Akt substrate 160, the most proximal step identified in the insulin signalling cascade regulating GLUT4 translocation and glucose uptake (Plomgaard *et al.* 2005).

With regard to IL-6, its role in insulin resistance is highly controversial (as reviewed in Pedersen & Febbraio, 2008). Infusion of recombinant human (rh) IL-6 into resting healthy humans does not impair whole body, lower limb, or subcutaneous adipose tissue glucose uptake or endogenous glucose production (EGP), although IL-6 contributes to the contraction-induced increase in endogenous glucose production. When diabetes patients were given rhIL-6-infusion, plasma concentrations of insulin declined to levels comparable with that in age- and BMI-matched healthy controls, indicating that the IL-6 enhanced insulin sensitivity.

A number of studies indicate that IL-6 enhances lipolysis, as well as fat oxidation, via an activation of AMPK (reviewed in Pedersen & Febbraio, 2008). Consistent with this idea, Wallenius *et al.* (2002) demonstrated that IL-6 deficient mice developed mature-onset obesity and insulin resistance. In addition, when the mice were treated with IL-6, there was a significant decrease in body fat mass in the IL-6 knock-out, but not in the wild-type, mice. To determine whether physiological concentrations of IL-6 affected lipid metabolism, our group administered physiological concentrations of rhIL-6 to healthy young and elderly humans as well as patients with type 2 diabetes (Pedersen *et al.* 2005; van Hall *et al.* 2003). The latter studies identified IL-6 as a potent modulator of fat metabolism in humans, increasing lipolysis as well as fat oxidation without causing hypertriacylglycerolaemia.

Of note, whereas it is known that both TNF- α and IL-6 induce lipolysis, only IL-6 appears to induce fat oxidation (van Hall *et al.* 2003; Plomgaard *et al.* 2007). Given the different biological profiles of TNF- α and IL-6 and given that TNF- α may trigger an IL-6 release, one theory holds that it is TNF- α derived from adipose tissue that is actually the major 'driver' behind inflammation-induced insulin resistance and atherosclerosis. Importantly, also tumour initiation, promotion and progression are stimulated by systemic elevation of pro-inflammatory cytokines (Lin & Karin, 2007; Handschin & Spiegelman, 2008).

A chronic inflammatory environment will lead to a state of insulin resistance with hyperinsulinaemia. The so-called hyperinsulinaemia hypothesis goes hand in hand with the inflammation hypothesis.

The hyperinsulinaemia hypothesis suggests that elevated levels of insulin and free IGF-1 promote proliferation of colon cells and lead to a survival benefit of transformed cells, ultimately resulting in colorectal cancer (Berster & Goke, 2008). In addition, a number of neurodegenerative diseases are linked to a local inflammatory response in the brain (neuroinflammation), e.g. interleukin-1 β (IL-1 β), TNF-related apoptosis-inducing ligand (TRAIL) and other cytokines have been postulated to be involved in the aetiology of Alzheimer's disease (Zipp & Aktas, 2006). Moreover, in addition to the neuroinflammation found in many neurodegenerative disorders, systemic inflammation may further exacerbate the progression of neurodegeneration (Perry *et al.* 2007).

The myokine concept

The protective effect of exercise against diseases associated with chronic inflammation may to some extent be ascribed to an anti-inflammatory effect of regular exercise. I hereby suggest that the long-term anti-inflammatory effects of

exercise may be mediated via effects of exercise leading to a reduction in visceral fat mass.

In line with the acceptance of adipose tissue as an endocrine organ, we came up with the innovative idea that also skeletal muscle should be viewed as an endocrine organ (Pedersen & Febbraio, 2008). In continuation, we have suggested that cytokines and other peptides that are produced, expressed and released by muscle fibres and exert paracrine or endocrine effects should be classified as 'myokines' (Pedersen & Febbraio, 2008). Given that skeletal muscle is the largest organ in the human body, our discovery of contracting muscle as a cytokine producing organ opens up a whole new paradigm: through evolution muscle has had a central role in orchestrating metabolism and function of other organs. This paradigm provides a conceptual basis explaining the multiple consequences of a physically inactive life style. If the endocrine function of the muscle is not stimulated through contractions, this will cause malfunction of several organs and tissues of the body as well as an increased risk of cardiovascular disease, cancer and dementia (Pedersen & Febbraio, 2008).

Today, it appears that skeletal muscle has the capacity to express several myokines. The list includes interleukin IL-6, IL-8, IL-15, brain-derived neurotrophic factor (BDNF) and leukaemia inhibitory factor (LIF) (Broholm *et al.* 2008). Through other strategies, Kenneth Walsh, Boston, has recently identified the myokines FGF21 and follistatin-like-1 (Izumiya *et al.* 2008; Ouchi *et al.* 2008). Thus, although the idea of an 'exercise factor' can be traced back many years, our recent identification of muscle as a myokine-producing organ opens up a whole new field of research. In this review, I briefly highlight some myokines with specific effect on fat oxidation.

IL-6: the myokine prototype. The first identified and most studied myokine is the gp130 receptor cytokine interleukin-6 (IL-6) (Pedersen & Febbraio, 2008). IL-6 was discovered as a myokine because of the observation that it increases up to 100-fold in the circulation during physical exercise. Identification of IL-6 production by skeletal muscle during physical activity generated renewed interest in the metabolic role of IL-6 because it created a paradox. On one hand, IL-6 is markedly produced and released in the post-exercise period when insulin action is enhanced, but on the other hand, IL-6 has also been associated with obesity and reduced insulin action (Pedersen & Febbraio, 2008). However, a number of studies during the past decade have revealed that in response to muscle contractions, both type I and type II muscle fibres express the myokine IL-6, which subsequently exerts its effects both locally within the muscle (e.g. through activation of AMPK) and – when released into the circulation – peripherally in several organs in a

hormone-like fashion. Within skeletal muscle, IL-6 acts locally to signal through a gp130R β /IL-6R α homodimer, resulting in activation of AMP kinase and/or PI3 kinase to increase glucose uptake and fat oxidation. IL-6 may also work in an endocrine fashion to increase hepatic glucose production during exercise or lipolysis in adipose tissue (reviewed in Pedersen & Febbraio, 2008). Although it has not been demonstrated that IL-6 has specific effects on visceral fat mass, it appears to play an important role in lipid metabolism as is illustrated in Fig. 3.

IL-15 – a role in muscle–fat cross talk? IL-15 is expressed in human skeletal muscle and has been identified as an anabolic factor in muscle growth. In addition to its anabolic effects on skeletal muscle *in vitro* and *in vivo*, IL-15 appears to play a role in lipid metabolism (Nielsen & Pedersen, 2007). Therefore, IL-15 was suggested to be involved in muscle–fat cross talk.

Recently, we demonstrated that IL-15 mRNA levels were upregulated in human skeletal muscle following a bout of strength training (Nielsen *et al.* 2007) suggesting that IL-15 may accumulate within the muscle as a consequence of

regular training. Interestingly, we further demonstrated a negative association between plasma IL-15 concentration and trunk fat mass, but not limb fat mass, in humans. In support of the human data, we demonstrated a decrease in visceral fat mass, but not subcutaneous fat mass, when IL-15 was overexpressed in murine muscle (Nielsen *et al.* 2008). Quinn *et al.* (2009) found that elevated circulating levels of IL-15 resulted in significant reductions in body fat and increased bone mineral content, without appreciably affecting lean body mass or levels of other cytokines. Although, the latter model represented an artificial system, the findings lend some support to the idea that IL-15 secretion from muscle tissue may modulate visceral fat mass in particular via an endocrine mechanism.

BDNF – a role in neurobiology and peripheral metabolism.

It was by realizing the disease cluster of physical inactivity that we recently became interested in brain-derived neurotrophic factor (BDNF), which is a member of the neurotrophic factor family. BDNF is recognized as playing a key role in regulating survival, growth and maintenance of neurons (Mattson *et al.* 2004), and it plays a role in learning and memory (Tyler *et al.* 2002). Hippocampal

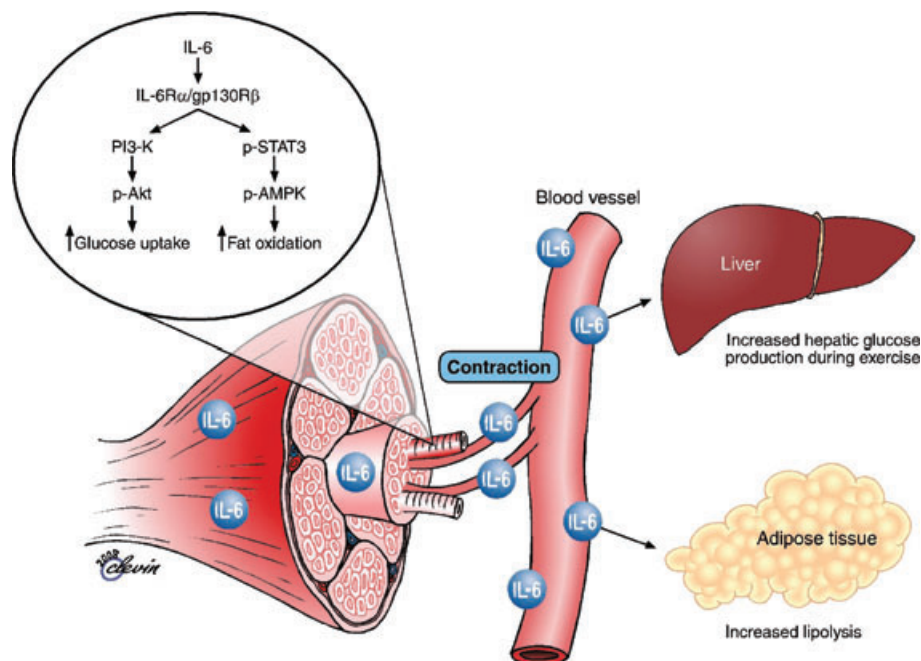


Figure 3. Biological role of contraction-induced IL-6

Skeletal muscle expresses and releases myokines into the circulation. In response to muscle contractions, both type I and type II muscle fibres express the myokine IL-6, which subsequently exerts its effects both locally within the muscle (e.g. through activation of AMPK) and – when released into the circulation – peripherally in several organs in a hormone-like fashion. Specifically, in skeletal muscle, IL-6 acts in an autocrine or paracrine manner to signal through a gp130R β /IL-6R α homodimer, resulting in activation of AMP kinase and/or PI3 kinase to increase glucose uptake and fat oxidation. IL-6 is also known to increase hepatic glucose production during exercise or lipolysis in adipose tissue. Modified from Pedersen & Febbraio (2008) with permission from the American Physiological Society and from Pedersen and Fischer (2007) with permission from Elsevier.

samples from Alzheimer's disease donors show decreased BDNF expression (Connor *et al.* 1997) and individuals with Alzheimer's disease have low plasma levels of BDNF (Laske *et al.* 2005). Also, patients with major depression have lower levels of serum BDNF than normal control subjects (Karege *et al.* 2002). Other studies suggest that plasma BDNF is a biomarker of impaired memory and general cognitive function in ageing women (Komulainen *et al.* 2008) and a low circulating BDNF level was recently shown to be an independent and robust biomarker of mortality risk in old women (Krabbe *et al.* 2009). In addition, it has previously been described that patients with acute coronary syndromes have reduced levels of BDNF in plasma (Manni *et al.* 2005). Interestingly, we found low levels of circulating BDNF also in individuals with both obesity and type 2 diabetes (Krabbe *et al.* 2007). Thus, BDNF is low in people with Alzheimer's disease, major depression, impaired cognitive function, CVD, type 2 diabetes and obesity, and constitutes a disease of low BDNF as illustrated in Fig. 4.

The finding of low plasma levels of BDNF in patients with type 2 diabetes stimulated us to study if the brain could potentially contribute to the systemic levels of BDNF. In a human *in vivo* model, we demonstrated that there is a cerebral output of BDNF at basal condition, and that cerebral output of BDNF is inhibited during hyperglycaemic clamp conditions in humans. The latter finding may explain the concomitant finding of low circulating levels of BDNF in individuals with type 2 diabetes, and the association between low plasma BDNF and the severity of insulin resistance (Krabbe *et al.* 2007). The latter human data are in accordance with reports from animal models suggesting that BDNF also plays a role in insulin resistance and in energy balance. Thus, BDNF reduces food intake and lowers blood glucose in genetically modified (*db/db*) obese mice; however, the hypoglycaemic effect of BDNF cannot be ascribed solely to the hypophagic effect of

BDNF, because BDNF administration has beneficial effects on glucose homeostasis and improves insulin resistance in *db/db* mice even when food-intake is controlled (Ono *et al.* 1997; Tonra *et al.* 1999; Nakagawa *et al.* 2000), suggesting that BDNF may also have peripheral metabolic effects.

We studied whether skeletal muscle would produce BDNF in response to exercise (Matthews *et al.* 2009). It was found that BDNF mRNA and protein expression were increased in human skeletal muscle after exercise; however, muscle-derived BDNF appeared not to be released into the circulation. BDNF mRNA and protein expression were increased in muscle cells that were electrically stimulated. Interestingly, BDNF increased phosphorylation of AMPK and Acetyl CoA Carboxylase (ACC) and enhanced fat oxidation both *in vitro* and *ex vivo*. Thus, we have been able to identify BDNF as a novel contraction-induced muscle cell-derived protein that may increase fat oxidation in skeletal muscle in an AMPK-dependent fashion, Fig. 2.

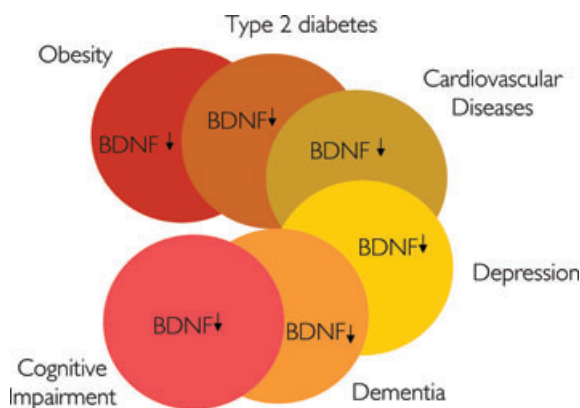


Figure 4. The disease of low circulating levels of brain derived neurotrophic factor (BDNF) has a major overlap with the diseases included in the disease of physical inactivity

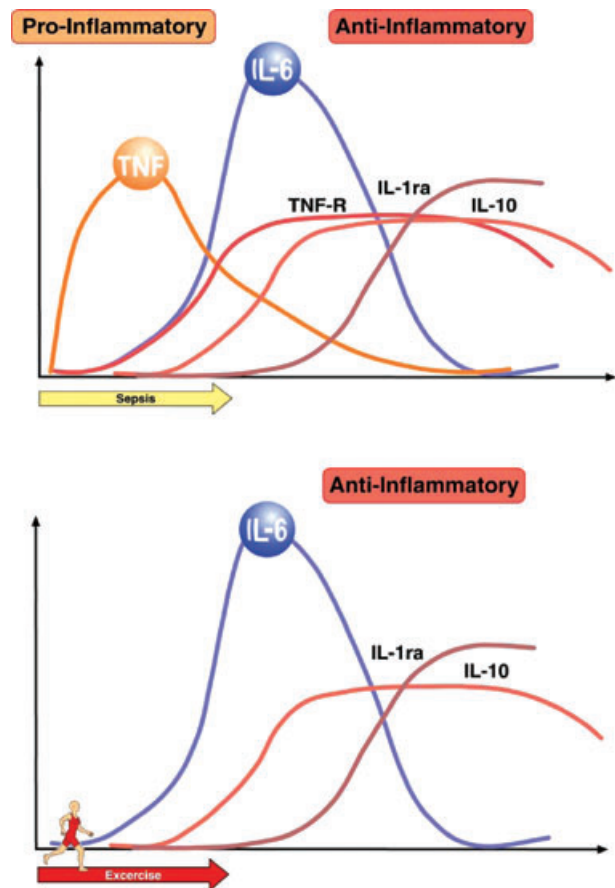


Figure 5. Comparison of sepsis-induced versus exercise-induced increases in circulating cytokines

During sepsis, there is a marked and rapid increase in circulating TNF- α , which is followed by an increase in IL-6. In contrast during exercise the marked increase in IL-6 is not preceded by elevated TNF- α . From Pedersen & Febbraio (2008) with permission from the American Physiological Society.

The possibility exists that BDNF may be classified as a myokine, which works in an autocrine or paracrine fashion with strong effects on peripheral metabolism, including fat oxidation with a subsequent effect on the size of adipose tissue.

Is EPO a new myokine? Most recent findings suggest that even erythropoietin (EPO) should be classified as a myokine (Hojman *et al.* 2009). Thus, we over-expressed EPO in murine skeletal muscles by gene electrotransfer, resulting in 100-fold increase in serum EPO and significant increases in haemoglobin levels. At 12 weeks, EPO expression resulted in a 23% weight reduction in EPO transfected obese mice. Correspondingly, dual energy x-ray absorptiometry (DXA) scanning revealed that this was due to a 28% reduction in adipose tissue mass. The decrease in adipose tissue mass was accompanied by a complete normalisation of fasting insulin levels and glucose tolerance in the high-fat fed mice. EPO expression also induced a 14% increase in muscle volume and a 25% increase in vascularisation of the EPO transfected muscle. In addition, muscular fat oxidation was increased 1.8-fold in both the EPO transfected and the contralateral muscle. Thus, although we were not able to draw conclusions with regard to the clinical relevance of these data, it appears that at least supra-physiological levels of EPO have substantial metabolic effects (Hojman *et al.* 2009). In the

latter study, EPO mimics a myokine in that it is produced and secreted from skeletal muscles and exhibits paracrine and endocrine effects on other muscles. However, the DNA electrotransfer method represents an artificial model to over-express and secrete EPO from muscles and our findings do not prove that EPO is indeed secreted natively from muscles. However, most recently Rundqvist *et al.* reported that EPO is released from the exercising leg to the circulation, possibly corresponding to an increased bioavailability of EPO. Although the release of EPO could only be verified in the initial period of an exercise bout, it suggests that EPO may be a true myokine (Rundqvist *et al.* 2009). Interestingly, the latter study reports that the EPO receptor (EPOR) mRNA and EPOR-associated Janus kinase (JAK) 2 phosphorylation were increased, suggesting that signalling through EPOR is involved in exercise-induced skeletal muscle adaptation.

Summary on Myokines. In the context of specific myokines, I suggest that physical inactivity is an *independent* cause of fat accumulation in 'the wrong places'. Accordingly, contracting skeletal muscles release myokines, which work in a hormone-like fashion, exerting specific endocrine effects on visceral fat and other ectopic fat deposits. Other myokines will work locally within the muscle via paracrine mechanisms, exerting their effects on signalling pathways involved in fat oxidation.

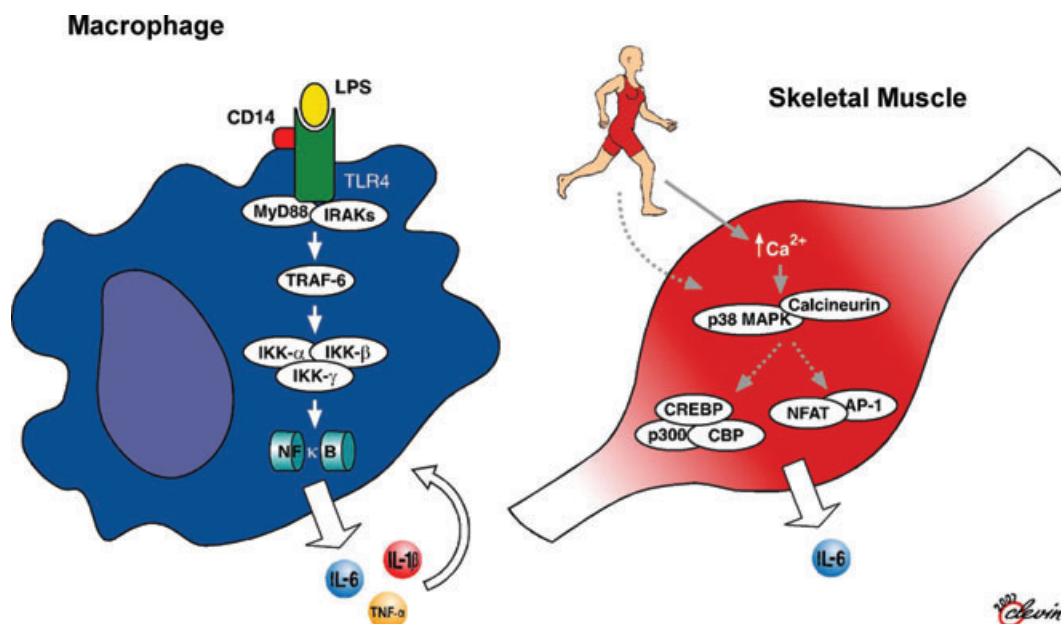


Figure 6. The proposed cytokine signalling pathways for macrophages and contracting skeletal muscle While it is well known that transcription of IL-6 and other pro-inflammatory cytokines such as TNF- α and IL- β is principally regulated by the TLR receptor signalling cascade that results in nuclear translocation and activation of NF- κ B, evidence in contracting skeletal muscle suggests that contraction leads to increased cytosolic Ca²⁺ and activation of p38 MAPK and/or calcineurin, which leads to activation of transcription factors depending upon these upstream events. From Pedersen & Febbraio (2008) with permission from the American Physiological Society.

The anti-inflammatory effects of an acute bout of exercise

Regular exercise appears to induce anti-inflammatory effects, suggesting that physical activity *per se* may suppress systemic low-grade inflammation. And several studies show that markers of inflammation are reduced following longer-term behavioural changes involving both reduced energy intake and increased physical activity (reviewed in Petersen & Pedersen, 2005). However, the mediators of this effect are unresolved. A number of mechanisms have been identified. Exercise increases the release of adrenaline, cortisol, growth hormone, prolactin and other factors that have immunomodulatory effects (Nieman, 2003; Handschin & Spiegelman, 2008).

To study whether acute exercise induces a true anti-inflammatory response, a model of 'low grade inflammation' was established in which a low dose of *E. coli* endotoxin was administered to healthy volunteers, who had been randomised to either rest or exercise prior to endotoxin administration. In resting subjects, endotoxin induced a 2- to 3-fold increase in circulating levels of TNF- α . In contrast, when the subjects performed 3 h of ergometer cycling and received the endotoxin bolus at 2.5 h, the TNF- α response was totally blunted (Starkie *et al.* 2003). This study provides some evidence that acute exercise may inhibit TNF production.

The cytokine response to exercise differs from that elicited by severe infections (see Fig. 5). The fact that the classical pro-inflammatory cytokines, TNF- α and IL-1 β , in general do not increase with exercise indicates that the cytokine cascade induced by exercise markedly differs from the cytokine cascade induced by infections (reviewed in Pedersen & Febbraio, 2008).

Typically, IL-6 is the first cytokine released into the circulation during exercise. The level of circulating IL-6 increases in an exponential fashion (up to 100-fold) in response to exercise and declines in the post-exercise period. The circulating levels of well-known anti-inflammatory cytokines such as IL-1ra and IL-10 also increase after exercise. Taken together, exercise provokes an increase primarily in IL-6, followed by an increase in IL-1ra and IL-10. The appearance of IL-6 in the circulation is by far the most marked and its appearance precedes that of the other cytokines, and a number of studies have demonstrated that contracting skeletal muscle fibres *per se* produce and release IL-6. Moreover, it appears that muscle-derived IL-6 may account for most of the systemic IL-6 response to exercise.

Recent work has shown that both up-stream and down-stream signalling pathways for IL-6 differ markedly between myocytes and macrophages (see also Fig. 6). It appears that unlike IL-6 signalling in macrophages, which is dependent upon activation of the NF κ B signalling pathway, intramuscular IL-6 expression is regulated by a network of signalling cascades that among other pathways

is likely to involve cross talk between the Ca²⁺ Nuclear Factor of activated T cells (NFAT) and glycogen/p38 MAPK pathways. Thus, when IL-6 is signalling in monocytes or macrophages, it creates a proinflammatory response, whereas IL-6 activation and signalling in muscle is totally independent of a preceding TNF response or NF κ B activation (Pedersen & Febbraio, 2008).

The possibility exists that with regular exercise, the anti-inflammatory effects of an acute bout of exercise will protect against chronic systemic low-grade inflammation, but such a direct link between the acute effects of exercise and the long-term benefits has yet to be established.

Conclusion and perspective

Much focus has been on the detrimental effects of abdominal adiposity. However, following adjustment for sex, age and hormonal levels, it remains a mystery why some people accumulate fat in 'the wrong places' (e.g. as visceral fat) and why others do not. In this review, physical inactivity is given the central role as an independent and strong risk factor for accumulation of visceral fat and other ectopic fat deposits. The finding that muscles produce and release myokines provides a conceptual basis for understanding some of the molecular mechanisms underlying organ cross talk, including muscle-fat cross talk. Accumulating data suggest that contracting skeletal muscles release myokines, which work in a hormone-like fashion, exerting specific endocrine effects on visceral fat and other ectopic fat deposits. Other myokines work locally within the muscle via paracrine mechanisms, exerting their effects on signalling pathways involved in fat oxidation. However, physical inactivity and muscle disuse lead to metabolic deterioration.

References

- Barabasi AL (2007). Network medicine – from obesity to the 'diseasome'. *N Engl J Med* **357**, 404–407.
- Bays HE (2009). 'Sick fat,' metabolic disease, and atherosclerosis. *Am J Med* **122**, S26–S37.
- Berster JM & Goke B (2008). Type 2 diabetes mellitus as risk factor for colorectal cancer. *Arch Physiol Biochem* **114**, 84–98.
- Broholm C, Mortensen OH, Nielsen S, Akerstrom T, Zankari A, Dahl B & Pedersen BK (2008). Exercise induces expression of leukaemia inhibitory factor in human skeletal muscle. *J Physiol* **586**, 2195–2201.
- Christakis NA & Fowler JH (2007). The spread of obesity in a large social network over 32 years. *N Engl J Med* **357**, 370–379.
- Connor B, Young D, Yan Q, Faull RL, Synek B & Dragunow M (1997). Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Brain Res Mol Brain Res* **49**, 71–81.
- Diamant M & Tushuizen ME (2006). The metabolic syndrome and endothelial dysfunction: common highway to type 2 diabetes and CVD. *Curr Diab Rep* **6**, 279–286.

- Festa A, D'Agostino R Jr, Tracy RP & Haffner SM (2002). Elevated levels of acute-phase proteins and plasminogen activator inhibitor-1 predict the development of type 2 diabetes: the insulin resistance atherosclerosis study. *Diabetes* **51**, 1131–1137.
- Fischer CP, Berntsen A, Perstrup LB, Eskildsen P & Pedersen BK (2007). Plasma levels of interleukin-6 and C-reactive protein are associated with physical inactivity independent of obesity. *Scand J Med Sci Sports* **17**, 580–587.
- Giovannucci E (2007). Metabolic syndrome, hyperinsulinemia, and colon cancer: a review. *Am J Clin Nutr* **86**, s836–s842.
- Haffner SM (2007). Abdominal adiposity and cardiometabolic risk: do we have all the answers? *Am J Med* **120**, S10–S16.
- Handschin C & Spiegelman BM (2008). The role of exercise and PGC1 α in inflammation and chronic disease. *Nature* **454**, 463–469.
- Hojman P, Brolin C, Gissel H, Brandt C, Zerahn B, Pedersen BK & Gehl J (2009). Erythropoietin over-expression protects against diet-induced obesity in mice through increased fat oxidation in muscles. *PLoS One* **4**, e5894.
- Hotamisligil GS (2003). Inflammatory pathways and insulin action. *Int J Obes Relat Metab Disord* **27**(Suppl 3), S53–S55.
- Izumiya Y, Bina HA, Ouchi N, Akasaki Y, Kharitonov A & Walsh K (2008). FGF21 is an Akt-regulated myokine. *FEBS Lett* **582**, 3805–3810.
- Karege F, Perret G, Bondolfi G, Schwald M, Bertschy G & Aubry JM (2002). Decreased serum brain-derived neurotrophic factor levels in major depressed patients. *Psychiatry Res* **109**, 143–148.
- Komulainen P, Pedersen M, Hanninen T, Bruunsgaard H, Lakka TA, Kivipelto M, Hassinen M, Rauramaa TH, Pedersen BK & Rauramaa R (2008). BDNF is a novel marker of cognitive function in ageing women: The DR's EXTRA Study. *Neurobiol Learn Mem* **90**, 596–603.
- Krabbe KS, Mortensen EL, Avlund K, Pedersen AN, Pedersen BK, Jorgensen T & Bruunsgaard H (2009). Brain-derived neurotrophic factor predicts mortality risk in older women. *J Am Geriatr Soc* **57**, 1447–1452.
- Krabbe KS, Nielsen AR, Krogh-Madsen R, Plomgaard P, Rasmussen P, Erikstrup C, Fischer CP, Lindegaard B, Petersen AM, Taudorf S, Secher NH, Pilegaard H, Bruunsgaard H & Pedersen BK (2007). Brain-derived neurotrophic factor (BDNF) and type 2 diabetes. *Diabetologia* **50**, 431–438.
- Laske C, Stransky E, Leyhe T, Eschweiler GW, Wittorf A, Richartz E, Bartels M, Buchkremer G & Schott K (2005). Stage-dependent BDNF serum concentrations in Alzheimer's disease. *J Neural Transm* **113**, 1217–1224.
- Lin WW & Karin M (2007). A cytokine-mediated link between innate immunity, inflammation, and cancer. *J Clin Invest* **117**, 1175–1183.
- Manni L, Nikolova V, Vyagova D, Chaldakov GN & Aloe L (2005). Reduced plasma levels of NGF and BDNF in patients with acute coronary syndromes. *Int J Cardiol* **102**, 169–171.
- Matter CM & Handschin C (2007). RANTES (regulated on activation, normal T cell expressed and secreted), inflammation, obesity, and the metabolic syndrome. *Circulation* **115**, 946–948.
- Matthews VB, Åström M-B, Chan MHS, Bruce CR, Prelovsek O, Åkerström T, Yfanti C, Broholm C, Mortensen OH, Penkowa M, Hojman P, Zankari A, Watt MJ, Pedersen BK & Febbraio MA (2009). Brain derived neurotrophic factor is produced by skeletal muscle cells in response to contraction and enhances fat oxidation via activation of AMPK. *Diabetologia* **52**, 1409–1418.
- Mattson MP, Maudsley S & Martin B (2004). BDNF and 5-HT: a dynamic duo in age-related neuronal plasticity and neurodegenerative disorders. *Trends Neurosci* **27**, 589–594.
- Monninkhof EM, Elias SG, Vlems FA, van der Tweel I, Schuit AJ, Voskuil DW & van Leeuwen FE (2007). Physical activity and breast cancer: a systematic review. *Epidemiology* **18**, 137–157.
- Nakagawa T, Tsuchida A, Itakura Y, Nonomura T, Ono M, Hirota F, Inoue T, Nakayama C, Taiji M & Noguchi H (2000). Brain-derived neurotrophic factor regulates glucose metabolism by modulating energy balance in diabetic mice. *Diabetes* **49**, 436–444.
- Nielsen AR, Hojman P, Erikstrup C, Fischer CP, Plomgaard P, Mounier R, Mortensen OH, Broholm C, Taudorf S, Krogh-Madsen R, Lindegaard B, Petersen AM, Gehl J & Pedersen BK (2008). Association between IL-15 and obesity: IL-15 as a potential regulator of fat mass. *J Clin Endocrinol Metab* **93**, 4486–4493.
- Nielsen AR, Mounier R, Plomgaard P, Mortensen OH, Penkowa M, Speerscheider T, Pilegaard H & Pedersen BK (2007). Expression of interleukin-15 in human skeletal muscle – effect of exercise and muscle fibre type composition. *J Physiol* **584**, 305–312.
- Nielsen AR & Pedersen BK (2007). The biological roles of exercise-induced cytokines: IL-6, IL-8, and IL-15. *Appl Physiol Nutr Metab* **32**, 833–839.
- Nieman DC (2003). Current perspective on exercise immunology. *Curr Sports Med Rep* **2**, 239–242.
- Nocon M, Hiemann T, Muller-Riemenschneider F, Thalau F, Roll S & Willich SN (2008). Association of physical activity with all-cause and cardiovascular mortality: a systematic review and meta-analysis. *Eur J Cardiovasc Prev Rehabil* **15**, 239–246.
- Olsen RH, Krogh-Madsen R, Thomsen C, Booth FW & Pedersen BK (2008). Metabolic responses to reduced daily steps in healthy nonexercising men. *JAMA* **299**, 1261–1263.
- Ono M, Ichihara J, Nonomura T, Itakura Y, Taiji M, Nakayama C & Noguchi H (1997). Brain-derived neurotrophic factor reduces blood glucose level in obese diabetic mice but not in normal mice. *Biochem Biophys Res Commun* **238**, 633–637.
- Ouchi N, Oshima Y, Ohashi K, Higuchi A, Ikegami C, Izumiya Y & Walsh K (2008). Follistatin-like 1, a secreted muscle protein, promotes endothelial cell function and revascularization in ischemic tissue through a nitric-oxide synthase-dependent mechanism. *J Biol Chem* **283**, 32802–32811.
- Paffenbarger RS Jr, Lee IM & Leung R (1994). Physical activity and personal characteristics associated with depression and suicide in American college men. *Acta Psychiatr Scand Suppl* **377**, 16–22.
- Pedersen BK (2007). Body mass index-independent effect of fitness and physical activity for all-cause mortality. *Scand J Med Sci Sports* **17**, 196–204.

- Pedersen BK & Febbraio MA (2008). Muscle as an endocrine organ: focus on muscle-derived interleukin-6. *Physiol Rev* **88**, 1379–1406.
- Perry VH, Cunningham C & Holmes C (2007). Systemic infections and inflammation affect chronic neurodegeneration. *Nat Rev Immunol* **7**, 161–167.
- Petersen AM & Pedersen BK (2005). The anti-inflammatory effect of exercise. *J Appl Physiol* **98**, 1154–1162.
- Petersen EW, Carey AL, Sacchetti M, Steinberg GR, Macaulay SL, Febbraio MA & Pedersen BK (2005). Acute IL-6 treatment increases fatty acid turnover in elderly humans *in vivo* and in tissue culture *in vitro*: evidence that IL-6 acts independently of lipolytic hormones. *Am J Physiol Endocrinol Metab* **288**, E155–E162.
- Pischon T, Boeing H, Hoffmann K, Bergmann M, Schulze MB, Overvad K, Van Der Schouw YT, Spencer E, Moons KGM, Tjonneland A, Halkjaer J, Jensen MK, Stegger J, Clavel-Chapelon F, Boutron-Ruault MC, Chajes V, Linseisen J, Kaaks R, Trichopoulos A, Trichopoulos D, Bamia C, Sieri S, Palli D, Tumino R, Vineis P, Panico S, Peeters PHM, May AM, Bueno-De-Mesquita HB, van Duijnhoven FJB, Hallmans G, Weinehall L, Manjer J, Hedblad B, Lund E, Agudo A, Arriola L, Barricarte A, Navarro C, Martinez C, Quiros JR, Key T, Bingham S, Khaw KT, Boffetta P, Jenab M, Ferrari P & Riboli E (2008). General and abdominal adiposity and risk of death in Europe. *New Engl J Med* **359**, 2105–2120.
- Plomgaard P, Fischer CP, Ibfelt T, Pedersen BK & van Hall G (2007). TNF- α modulates human *in vivo* lipolysis. *J Clin Endocrinol Metab* **93**, 543–549.
- Plomgaard P, Keller P, Keller C & Pedersen BK (2005). TNF- α , but not IL-6, stimulates plasminogen activator inhibitor-1 expression in human subcutaneous adipose tissue. *J Appl Physiol* **98**, 2019–2023.
- Quinn LS, Anderson BG, Strait-Bodey L, Stroud AM & Argiles JM (2009). Oversecretion of interleukin-15 from skeletal muscle reduces adiposity. *Am J Physiol Endocrinol Metab* **296**, E191–E202.
- Richardson LC & Pollack LA (2005). Therapy insight: Influence of type 2 diabetes on the development, treatment and outcomes of cancer. *Nat Clin Pract Oncol* **2**, 48–53.
- Rovio S, Kareholt I, Helkala EL, Viitanen M, Winblad B, Tuomilehto J, Soininen H, Nissinen A & Kivipelto M (2005). Leisure-time physical activity at midlife and the risk of dementia and Alzheimer's disease. *Lancet Neurol* **4**, 705–711.
- Rundqvist H, Rullman E, Sundberg CJ, Fischer H, Eisleitner K, Stahlberg M, Sundblad P, Jansson E & Gustafsson T (2009). Activation of the erythropoietin receptor in human skeletal muscle. *Eur J Endocrinol* **161**, 427–434.
- Starkie R, Ostrowski SR, Jauffred S, Febbraio M & Pedersen BK (2003). Exercise and IL-6 infusion inhibit endotoxin-induced TNF- α production in humans. *FASEB J* **17**, 884–886.
- Tonra JR, Ono M, Liu X, Garcia K, Jackson C, Yancopoulos GD, Wiegand SJ & Wong V (1999). Brain-derived neurotrophic factor improves blood glucose control and alleviates fasting hyperglycemia in C57BLKS-Lepr(db)/lepr(db) mice. *Diabetes* **48**, 588–594.
- Tuomilehto J, Lindstrom J, Eriksson JG, Valle TT, Hamalainen H, Ilanne-Parikka P, Keinanen-Kiukaanniemi S, Laakso M, Louheranta A, Rastas M, Salminen V & Uusitupa M (2001). Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med* **344**, 1343–1350.
- Tyler WJ, Alonso M, Bramham CR & Pozzo-Miller LD (2002). From acquisition to consolidation: on the role of brain-derived neurotrophic factor signalling in hippocampal-dependent learning. *Learn Mem* **9**, 224–237.
- van Hall G, Steensberg A, Sacchetti M, Fischer C, Keller C, Schjerling P, Hiscock N, Moller K, Saltin B, Febbraio MA & Pedersen BK (2003). Interleukin-6 stimulates lipolysis and fat oxidation in humans. *J Clin Endocrinol Metab* **88**, 3005–3010.
- Wallenius V, Wallenius K, Ahren B, Rudling M, Carlsten H, Dickson SL, Ohlsson C & Jansson JO (2002). Interleukin-6-deficient mice develop mature-onset obesity. *Nat Med* **8**, 75–79.
- Whitmer RA, Gustafson DR, Barrett-Connor E, Haan MN, Gunderson EP & Yaffe K (2008). Central obesity and increased risk of dementia more than three decades later. *Neurology* **71**, 1057–1064.
- Wolin KY, Yan Y, Colditz GA & Lee IM (2009). Physical activity and colon cancer prevention: a meta-analysis. *Br J Cancer* **100**, 611–616.
- Xue F & Michels KB (2007). Diabetes, metabolic syndrome, and breast cancer: a review of the current evidence. *Am J Clin Nutr* **86**, s823–s835.
- Yudkin JS (2007). Inflammation, obesity, and the metabolic syndrome. *Horm Metab Res* **39**, 707–709.
- Yudkin JS, Eringa E & Stehouwer CD (2005). 'Vasocrine' signalling from perivascular fat: a mechanism linking insulin resistance to vascular disease. *Lancet* **365**, 1817–1820.
- Zipp F & Aktas O (2006). The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends Neurosci* **29**, 518–527.

Acknowledgements

I wish to gratefully acknowledge my collaborators, post-doctoral fellows, students and technicians who have contributed much of the work reported in this review. The Centre of Inflammation and Metabolism is supported by a grant from the Danish National Research Foundation (DG 02-512-555). In addition, support was obtained from the Danish Medical Research Council and the Commission of the European Communities (contract no. LSHM-CT-2004-005272 EXGENESIS). The Capital Region of Copenhagen and the Copenhagen University support the Copenhagen Muscle Research Centre.